

780 Dubuque Avenue
So. San Francisco, CA 94080, U.S.A.
Tel: (800) 989-6296 / Fax:(650)871-2857
<http://www.maximbio.com>
E-mail: mbi@maximbio.com

MPCR Kit for Mouse TNF Signaling Genes Set-3
Cat No. MP-70212: 50 reactions
Cat No. MP-70211: 100 reactions

**INSTRUCTION
MANUAL**

Revised March 4, 2003

*These products are designed and sold for use in the Multiplex PCR (MPCR) covered by patent # 5,582,989. Use of the MPCR process requires a license. A limited, non-automated research field license under the patent to use only this amount of the product to practice the MPCR process is conveyed to the purchaser by the purchase of this product.

The Polymerase Chain Reaction (PCR) process is covered by patents owned by Hoffman-LaRoche. Use of the PCR process requires a license. A license for diagnostic purposes may be obtained from Roche Molecular System. A license for research may be obtained by the purchase and the use of authorized reagents and DNA thermocyclers from the Perkin-Elmer Corporation or by negotiating a license with Perkin-Elmer.

This product is intended for research use only and not for diagnostic purposes.

INTRODUCTION

Mammalian cells can self-destruct via intrinsic programmed cell death (1). Apoptosis, a form of programmed cell death, is characterized by specific morphologic and biochemical properties. Apoptosis plays a major role in many biological processes, including embryogenesis, development of the immune system and tissue regeneration. Like growth and differentiation, apoptosis requires active and coordinated regulation of specific genes. Some molecular components of the apoptotic program have been conserved through evolution. Genetic studies of *C. elegans* have led to the identification of mutations, on 14 genes, that affect programmed cell death in this organism (1). Two of these genes, *ced-9* and *ced-3* are homologous to mammalian genes: the proto-oncogene *bcl-2* and ICE (interleukin-1- β converting enzyme) respectively.

Ecotopic expression of *bcl-2* has been shown to block apoptosis in many experimental systems (2). Although the mechanism of *bcl-2* action is unknown, biochemical studies have implicated this protein in the regulation of cell redox potential. Genetic evidence indicates that *bcl-2* belongs to an emerging family. Some members of the *bcl-2* family, like *bcl-xL* (3), suppress apoptosis while other members, such as *bax* and *bcl-xS* (3,4), increase the susceptibility of cells to apoptotic stimuli. In addition, *bcl-2* family members form hetero- and homo-dimers, suggesting that cellular susceptibility to apoptosis is partly influenced by the level of gene expression.

ICE encodes a cysteine protease that cleaves peptide bonds after Asp residues (5). A family of ICE genes (*Ich-1/Nedd-2*, *cpp32b/yama*, *Tx/Ich-2*, and *Mch-2*) have been identified and appear to induce apoptosis when they are overexpressed in various cell types (5). ICE inhibitors, such as the cowpox viral protein CrmA and the peptide YVAD, block apoptosis when it is induced by Fas ligand or Fas receptor cross-linking.

P53, a DNA binding protein and transcriptional activator, regulates cell proliferation and induces apoptosis in certain circumstances and cellular backgrounds. Studies have clearly shown that when P53 minus cell lines are transfected with a temperature sensitive mutant of P53, the wild-type P53 lowers the threshold for inducing apoptosis following genotoxic damage(6). Thus, P53 may function as a sensor for DNA damage.

The *myc* oncogene has been intensively studied for its roles in both cell proliferation and apoptosis since *myc* is known to be a transcriptional regulator. It has been found that levels of *myc* expression correlate with proliferative capacity and susceptibility to apoptosis (7).

Analysis of the temporal and spatial distribution of RNA expression provides researchers with important clues about the function of apoptosis regulating genes in their own systems. Northern Blot and RNase Protection Assay are the most widely used procedures for determining the abundance of a specific mRNA in a total or poly(A) RNA sample. RT-MPCR provides an alternate and accurate method to detect multiple gene expression by amplifying all the genes under the same conditions (8a,8b & 8c). Variations in RNA isolation, initial quantitation errors or tube-to-tube variations in RT and PCR can be compensated by including a house-keeping gene, such as GAPDH, in MPCR. Alternatively, a parallel RT-PCR using the same cDNA, PCR conditions and primers for one of house-keeping genes may be run to offset any variations. Differences in gene expression can be determined by normalizing its expression against GAPDH expression.

Maxim's mTNF3G-MPCR kits have been designed to detect the expression of mouse GAPDH, Cox-2, NFkB, c-myc, Fas, TNF-a, *bcl-2*, P53 and Ikb genes. The PCR primers have similar Tm and no obvious 3'-end overlap to enhance multiple amplification. The 658 bp(GAPDH), 449 bp (Cox-2), 409 bp(NFkB), 348 bp (c-myc), 316 bp(Fas), 274 bp(TNF-a), 235 bp(*bcl-2*), 205 bp (P53) and 189 bp(Ikb) PCR products can be generated from mouse RNA or the positive control, which is included in this kit. Therefore, the mTNF-MPCR kit provides a quick and simple method to analyze Cox-2, NFkB, c-myc, Fas, TNF-a, *bcl-2*, P53 and Ikb gene expression and normalize their expression against GAPDH gene expression.

PCR PRODUCT QUANTITATION

I: Radioactive Quantitation

In our experience, visual inspection of an EtBr-stained agarose gel is sensitive and precise enough to detect changes as low as two-fold. If greater discrimination is necessary, several methods are available. The simplest procedure is to add a radioactively labeled dNTP to the PCR reaction. After gel analysis, the band may be excised and counted in a scintillation counter. Alternatively the gel may be dried and an autoradio-gram may be generated which can be scanned in a densitometer. Another method is to label the 5' end of one or both of the primers with ^{32}P , which is incorporated into the PCR products and then assayed for radioactivity (13).

Southern blot hybridization with synthetic DNA probes may also be performed to verify and quantitate PCR generated products, either by densitometry of an autoradiogram or by excising and counting the signal from a hybridization membrane. This method also quantitates only the target product without interference from nontarget products or primer-generated artifacts.

II: Non-Radioactive Quantitation

Nonradioactive quantitation methods include the use of biotinylated or digoxigenin-labeled primers in conjunction with the appropriate detection methods (14), use of a bioanalyzer or WAVE. For an in-depth discussion of the various methods of PCR product quantitation, refer to the review article by Bloch (15).

In addition to the above methods, several companies now offer gel video systems which can scan and quantitate EtBr-stained gel bands in much the same way a densitometer does. Lab-on-a-chip (BioAnalyzer), CE, HPLC, and WAVE may also be used to analyze MPCR products and quantitate simultaneously.

COMPARISON OF MPCR WITH RPA

MPCR (Multiplex Polymerase Chain Reaction)

- √ Non-isotope method with high sensitivity
0.1-1 μg total RNA per MPCR
- √ Whole process takes only a few hours
- √ Detect Multiple Genes Simultaneously & Quantitatively
- √ Signal can be quantified directly from gel if isotope is included in MPCR. Additional techniques can be used to quantify MPCR product (using Bioanalyzer, HPLC, and WAVE.)
- √ Non-specific products can be eliminated by using probes and southern hybridization.
- √ Ready-to-use

RPA (RNase Protection Assay)

- √ Isotope or Non-Isotope methods
1-20 μg total RNA per RPA assay
- √ Whole process takes two days
- √ Detect Multiple Genes Simultaneously & Quantitatively
- √ Signal can be quantified directly from gel
- √ Non-specific signal can be generated by either low stringent conditions or high-secondary-structure template.
- √ Make own "hot" RNA probes

MPCR KIT DESCRIPTION

MPCR Amplification Kits include all necessary MPCR amplification reagents with the exception of *Taq* Polymerase. These kits have been designed to direct the simultaneous amplification of specific regions of human DNA.

MPCR Kits come in two quantities:

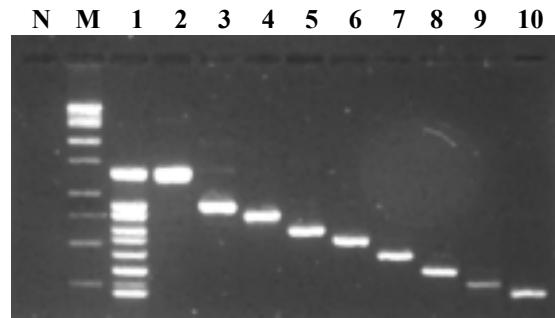
- 50X 50 μ L reaction kits
- 100X 50 μ L reaction kits

Each kit offers Maxim's optimal primer/buffer system which will enhance amplification specificity.

Figure 1 shows quality control MPCR results obtained by following MPCR kit manual using different concentrations of positive control.

For optimal results, please read and follow the instructions in this manual carefully. If you have any questions, please contact Maxim Biotech Customer Service at (650) 871-1919.

Figure 1



Lane N: PCR using mTNF3G Primers without positive control (Negative)
 Lane 1: PCR using mTNF3G Primers with positive control
 Lane 2: PCR using Mouse GAPDH Primers with positive control
 Lane 3: PCR using Mouse Cox-2 Primers with positive control
 Lane 4: PCR using Mouse NFK-B Primers with positive control
 Lane 5: PCR using Mouse c-myc Primers with positive control
 Lane 6: PCR using Mouse Fas Primers with positive control
 Lane 7: PCR using Mouse TNF-a Primers with positive control
 Lane 8: PCR using Mouse bcl-2 Primers with positive control
 Lane 9: PCR using Mouse P53 Primers with positive control
 Lane 10: PCR using Mouse Ik-B Primers with positive control
 M: DNA M.W. Marker

MPCR PRIMER INFORMATION

Product Code	Gene	5'/3' Tm	Amplicon Size	Accession No.	Intron Span	Genomic Size
mTNF3G-IkB	Mouse Ik-B	67°C/71°C	189bp	U19799	no	189bp
mTNF3G-P53	Mouse P53	69°C/70°C	205bp	M13874	yes	684bp
mTNF3G-BCL2	Mouse bcl-2	70°C/71°C	235bp	M16506	no	235bp
mTNF3G-TNF	Mouse TNF-a	69°C/70°C	274bp	M13049	yes	1260bp
mTNF3G-FAS	Mouse Fas	65°C/68°C	316bp	M83649	yes	9788bp
mTNF3G-MYC	Mouse c-myc	68°C/69°C	348bp	X01023	no	348bp
mTNF3G-NFKB	Mouse NFK-B	67°C/69°C	409bp	M61909	yes	2359bp
mTNF3G-Cox-2	Mouse Cox-2	68°C/68°C	449bp	NM_011198	no	449bp
mTNF3G-GAP	Mouse GAPDH	68°C/69°C	658bp	M32599	no	658bp

KIT COMPONENTS

MP-70212

50X50 μ L MPCR reaction kit
Store all reagents at -20°C

Product Code	Kit Component	Amount
mTNF3G -B001	2X mTNF3G MPCR Buffer (containing chemicals, enhancer, stabilizer and dNTPs)	1250 μ l
mTNF3G -C002	10X mTNF3G MPCR Pos. Control	50 μ l
mTNF3G -P002	10X mTNF3G MPCR Primers	250 μ l
MRB-0014	DNA M.W. Marker (100bp Ladder)	100 μ l
MRB-0011P	ddH ₂ O (DNase free)	2.0 ml
	Instruction Manual	

MP-70211

100X50 μ L MPCR reaction kit
Store all reagents at -20°C

Product Code	Kit Component	Amount
mTNF3G-B001	2X mTNF3G MPCR Buffer (containing chemicals, enhancer, stabilizer and dNTPs)	1250 μ l X2
mTNF3G-C002	10X mTNF3G MPCR Pos. Control	50 μ l X2
mTNF3G-P002	10X mTNF3G MPCR Primers	250 μ l X2
MRB-0014	DNA M.W. Marker (100bp Ladder)	100 μ l X2
MRB-0011P	ddH ₂ O (DNase free)	2.0 ml X2
	Instruction Manual	

NOTE: SPIN ALL TUBES BEFORE USING AND VORTEX ALL REAGENTS FOR AT LEAST 15 SECONDS BEFORE USING!!

PROCEDURE

RT Protocol:

The isolation of undegraded, intact RNA is an essential prerequisite for successful first strand synthesis and PCR amplification. Care should be taken to avoid RNase contamination of buffers and containers used for RNA work by pretreating with DEPC, autoclaving, and baking. Always wear sterile gloves when handling reagents. Use cDNA derived from 10^5 cells ($1\mu\text{g}$ cDNA) and apply them to each MPCR reaction.

1. Prepare total RNA, mRNA or use the control GAPDH RNA which is provided in Maxim's MPCR kit. **NOTE:** It is best to use cDNA derived from $0.5\text{-}1 \times 10^5$ cells ($0.5\text{-}1\mu\text{g}$ cDNA derived from RNA) for each MPCR reaction.
2. Equilibrate 3 water baths: 37°C , 70°C and 95°C .
3. **On ice**, pipet $1\text{-}2 \mu\text{g}$ mRNA or $10 \mu\text{g}$ total RNA (from 10^6 cells) dissolved in pure water or $2 \mu\text{l}$ control GAPDH RNA into a RNAase free reaction vial. We strongly recommend including a positive control reaction when setting up an RT-PCR reaction for the first time.
4. Add sterile water to a final volume of $14.5 \mu\text{l}$.
5. Add $4 \mu\text{l}$ random hexamer (50 mM) or Oligo(dT) (50 mM).
NOTE: The hexamer and Oligo(dT) RT reactions may be run simultaneously.
6. Incubate tube(s) at 70°C for 5 minutes and quickly chill on ice.
7. Begin your RT reaction by adding the following reagents to your hexamer or Oligo mixture:

Reagent	Description	Volume per Reaction
RNase Inhibitor	130 U/ μl	$0.5 \mu\text{l}$
5 X RT buffer	250mM Tris-HCl (pH8.3) 375mM KCl, 15mM MgCl_2 , 50mM DTT	$10 \mu\text{l}$
dNTPs	1mM each	$20 \mu\text{l}$
MMLV RT	250U/ μl	$1 \mu\text{l}$

8. Incubate the RT mixture at 37°C for 60 minutes.
9. Then, heat RT mixture at 95°C for 10 minutes and quickly chill on ice. This will help to eliminate the RT enzyme interference of MPCR reaction later.
10. Add another **$50 \mu\text{l}$** water or 0.1X TE buffer.
11. **$2\text{-}5 \mu\text{l}$** of above cDNA is sufficient for most genes in a standard MPCR reaction. However, more or less DNA may be needed in PCR depending on the copy number of the specific gene.

NOTE: Please do not use excess amount of cDNA. The salt from RT reaction may interfere the performance of MPCR.

PCR Protocol:

1. *Taq* DNA polymerase from Perkin-Elmer or its derivatives are highly recommended for MPCR. Ampli-*Taq* Gold, however, is not recommended because its own optimal buffer system is required.
2. **Reaction Mixture Preparation:**
 - A. Set up MPCR reactions with the test samples and MPCR buffers provided in the MPCR kit according to the table on the next page:

PROCEDURE

Volume (Per assay)	Reagent (Add in order)
25.0 µl	2X MPCR BufferMixture
5.0 µl	10X MPCR Primers
0.5 µl	<i>Taq</i> DNA Polymerase(5U/µl)
5.0 µl	Specimen cDNA or 10X Control cDNA from kit
14.5 µl	H ₂ O
50.0 µl	Mineral Oil (optional)

*: ³²P dNTPs may be used here to achieve higher sensitivity and better quantitation. 5-10 µCi [³²P]dCTP (3000 Ci/mmmole) should be used here per MPCR. Keep final dNTPs concentration same as without ³²P-dNTPs.

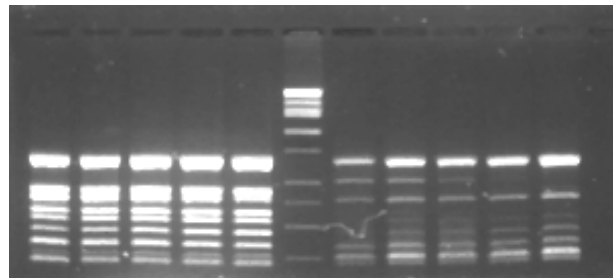
B. EDTA concentration in test sample must not exceed 0.5 mM because Mg⁺⁺ concentration in MPCR Buffers is limited to certain ranges. Additional Mg⁺⁺ may be added to the PCR mixture to compensate for EDTA. We strongly recommend running an MPCR reaction with the positive control provided in the kit. Since the MPCR DNA polymerase needed in each reaction is in a very small volume, it is recommended that all of the PCR components be premixed in a sufficient quantity for daily needs and then dispensed into individual reaction vials. This will help you to achieve more accurate measurements.

3. PCR thermocycle profile:

Reaction profiles will need to be optimized according to the machine type and needs of user. Please take note that temperature variations occur between different thermocyclers, therefore, the annealing temperature in the sample profile below is given as a range. It will be necessary to determine the optimal temperature for your individual thermocycler. An example of a time-temperature profile for the positive control PCR reaction optimized for Perkin Elmer machine types 480, 2400, and 9600 is provided below:

Temperature	Time	Cycles
96°C 58-60°C*	1 min 4 min	2X
94°C 58-60°C*	1 min 2 min	28-35X
70°C	10 min	1X
25°C	soak	

55 58 61 64 67°C



*The performance of MPCR kit against annealing temperatures. The above gel picture is an illustration of different annealing temperatures on MPCR kit MP-70211.

Note: A 2-step PCR thermocycle profile was found to be more effective than a 3-step PCR thermocycle profile for MPCR amplification. For 2-step PCR, use 94-95°C for denaturation and 58-60°C for annealing and extension. The 72°C step is omitted.

4. Agarose Gel Electrophoresis:

To fractionate the MPCR DNA product electrophoretically, mix 10µl of the MPCR product with 2µl 6X loading buffer. Run the total 12µl alongside 10 µl of DNA marker* from the MPCR kit on a 2 % agarose gel containing 0.5 mg/ml ethidium bromide. Electrophorese and photograph. (Hint: Best results are obtained when the gels are run slowly at less than 100 volts).

* DAN Marker contains linear double stranded DNA bands of 1,000; 900, 800, 700; 600; 500; 400; 300; 200; and 100 base pairs (bp).

TROUBLESHOOTING

1. MPCR AMPLIFICATION

Observation	Possible Cause	Recommended Action
1.1. No signal or missing some bands during amplification even using positive control provided in kit.	1.1a. The annealing temperature in the thermocycler is too high. 1.1b. Dominant primer dimers.	1.1a. Decrease PCR annealing temperature 3-5°C gradually. 1.1b. Use any one of "Hot Start" PCR procedures.
1.2. Too many nonspecific bands.	1.2a. The annealing temperature in the thermocycler is too low. 1.2b. Pre-PCR mispriming. 1.2c. cDNA is interfering with MPCR	1.2a. Increase PCR annealing temperature 3-5°C gradually. 1.2b. Use any one of "Hot Start" PCR procedures. 1.2c. Clean cDNA with Phenol/ Chloroform. 1.2d. Use Maxim's 3M™-MPCR Kit.
1.3. No difference in gene expression among treatments	1.3a. PCR amplification of this specific gene has passed the exponential phase. 1.3b. Variation in sample preparation, RT reaction and amounts of input cDNA.	1.3a. Decrease PCR cycle number or decrease the input cDNA. 1.3b. Run a parallel PCR with a house-keeping gene to eliminate variables.

PRECAUTIONS AND STORAGE

Storage

1. Store all MPCR Kit components at -20°C. Under these conditions components of the kit are stable for 1 year.
2. Isolate the kits from any sources of contaminating DNA, especially amplified PCR product.
3. Do not mix MPCR kit components that are from different lots. Each lot is optimized individually.

REFERENCES

1. Stellar, H. (1995) *Science* 267, 1445-1449.
2. Reed, J.C. (1994) *J.Cell Biol.* 124, 1-6.
3. Boise, L.H. (1993) *Cell* 74, 597-608.
4. Oltvai, Z.N. et. al., (1993) *Cell* 74, 609-619.
5. Martin, S.J. and Green, D.R. (1995) *Cell* 82, 349-352.
6. Lowe, S.W., et. al., (1993) *Nature* 362, 847-849.
7. Amati, A. et.al., (1993) *EMBO J.* 112, 5083-5087.
- 8a. Maxim Biotech Tools, 1, 2-5, 1995.
- 8b. Chamberlain, J.S. et al., In: *The polymerase chain reaction*. Mullis K, Ferre F and Gibbs R, eds. Birkhauser Boston Press, 38-46, 1994.
- 8c. Kumar, A. et al., (1997) *Science* 278, 1630-1632.
9. Chumakov, K.M. (1994), RT can inhibit PCR and stimulate primer-dimer formation. *PCR Methods and Applications*. 4: 62-64.
10. Hayashi, K., Orita, M., Suzuki, Y. & Sekiya, T. (1989) *Nucleic Acids Res.* 17:3605.
11. Landgraf, A., Reckmann, B., & Pingoud, A. (1991) *Analytical Biochemistry* 193:231.
12. Bloch, W. (1991) *Biochemistry* 30:2735.